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# Morphological Indices and Carcass Measurements of Indigenous Breeds of Rams Intensively Fattened

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**Abstract:** Growth rate assessment is an important husbandry practice often carried out in livestock breeding and fattening. Meat production is the most important trait in the breeding objectives of sheep production. Useful information on the suitability of the animals for meat production could be evaluated on a morphological basis. The aim of this study was to investigate breed differences in liveweight, carcass traits and muscle distribution for meat characteristics among Balami (BA), Uda (UD), Yankasa (YK) and West African Dwarf rams (WAD). A total of forty-eight rams were randomly assigned into four groups of twelve rams per breed in a completely randomized design. Body linear measurements were obtained weekly during the fattening period. At the end of the feeding trial, animals were subjected to an 18-hours fast and slaughtered for carcass measurements and muscle distribution. Results showed that BA and UD had higher values for body length (115.75±4.25 and 110.25±1.89cm), withers height (70.50±5.57 and 69.00±2.16cm) and heartgirth (92.00±2.94 and 84.00±1.41cm), than YK (96.50±3.87, 60.75±3.30 and 76.75±0.96cm), respectively which in turn had higher values than WAD (86.50±5.20, 53.25±5.32 and 75.25±3.30cm). Carcass length, pistol length, and leg length were 75.75±4.79, 60.75±7.14 and 88.00±2.16; 75.50±4.66, 56.00±5.00 and 80.25±5.80; 59.00±7.79, 56.00±10.62 and 71.50±2.65; 60.50±3.11, 53.75±5.91 and 61.00±2.94cm for BA, UD, YK and WAD rams, respectively. BA and UD had higher proportion of muscle distribution and were generally better than the YK and WAD breeds. This may boost their potential for large scale meat production under fattening conditions.

**Keywords:** Hindquarter, Meat, Fattening, Carcass, Pistola, Muscle Distribution

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## 1. Introduction

Nigeria's livestock industry is small and slow-growing relative to the population relying on it for meat [1]. Farmers need to invest in breeds that have traits for good carcass quality. The breed that can deliver good quality meat should be selected or bred for important and desirable traits. The commercial value of meat animals is determined by the carcass weight, proportion, and distribution of carcass muscle [2]. The use of specialized meat breeds to improve animal productivity and economic results is desirable in intensive

sheep production. The quality of rams of local breeds is an important instrument to promote the consumption of meat in the local population [3]. Morphometric measurements have been used to evaluate the characteristics of various breeds of animals and could provide first-hand information on the suitability of animals for selection [4]. Based on body conformation, meat production can better be estimated than other production properties, making body measurements important selection criteria [5]. Muscularity indicates the amount of muscle tissue, determined by muscle to bone ratio and fatness describes the external and internal deposition of

fat in the carcass. According to Srivastava *et al.* [6], body measurements are indices of skeletal development and indirectly help to determine carcass composition. Another parameter that has been used over the years to estimate the amount of meat that could be obtained from slaughtered animals is carcass measurement. Carcass composition assessment assigns value, sorting of carcasses for further processing and transfers information back to the production sector, ensuring that carcasses meet consumer demand. The objectives of the present study were to analyze the interrelationships between weights and measurements of carcasses; compare various measurements of the body, carcass, and tissue distribution for BA, UD, YK and WAD breeds of ram under intensive management.

## 2. Materials and Methods

### 2.1. Location of Experiment

The study was conducted at the Sheep and Goat unit of the Teaching and Research Farm, University of Ibadan, located on Latitude 7°26'N and 3°54'E and on an elevation of 330m above sea level. The area is characterized by humid tropical climate with an annual rainfall of 1540 mm and an average temperature of 22.1°C to 30.8°C for ten-hours per day

### 2.2. Animal Management

Forty-eight rams of Balami (BA), Uda (UD), Yankasa (YK) and West African Dwarf (WAD) breeds. Weighing 17.00±0.39–19.38±0.39 (12-18months of age). The age of the animals was estimated by visual observation, interviewing the owners and by dentition. Four breeds of rams (Twelve replicate per breed) were raised intensively. The rams were dewormed (300mg albendazole bolus for internal and with acarimic spray for external parasites, respectively), dipped and vaccinated against known parasites and diseases during the period of twenty-one days and were adapted for 15 days followed by ninety days feeding period. Experimental animals were fattened for ninety days, during which time each pen of rams was supplied with concentrate (14% crude protein and 2.00 Mcal/kg digestible energy) and hay at the rate 5% (60:40) of the body weight per ram and *adlibitum* access to salt lick and fresh, clean and cool water. At the end of the ninety days, four rams in each group whose weights were the closest to the average final weight of the rams of respective breeds were chosen for subsequent carcass analysis.

### 2.3. Linear Body Measurements

The six morphometric traits measured were wither height, pelvis height, body length, heart girth, chest width, and chest depth. Measurement was done using a graduated measuring stick. To achieve this, animals were placed on flat ground and held by two field assistants. The length measurements were done using a Dalton's weighing band tape rule while the width measurements were taken using a calibrated wooden caliper [7]. The following body measurements were

estimated:

- 1) Body length: the distance between the point of shoulder and the pin-bone.
- 2) Height at withers: vertical distance from the withers to the floor.
- 3) Heart girth: circumference of the body just behind the fore legs
- 4) Chest depth: vertical distance from the withers to the chest bottom,
- 5) Chest width: width of the body at the withers.
- 6) Height at Pelvic: hind leg length between malleolus and perineum

### 2.4. Slaughter Procedure

The selected rams (four rams per breed) were transported to the slaughter house of the Department of Animal Science, University of Ibadan where pre-slaughter liveweight was recorded after twelve-hours of fasting but free access to water. Exsanguination was carried out by an incision on the jugular furrow at the occipital-atlantal junction close to the head, severing the carotid arteries, jugular veins, trachea, oesophagus, and the spinal cord. The head was removed and the carcass was hung by the hindlegs using a pulley and then skinned [8].

### 2.5. The Evisceration of Carcasses and Carcass Evaluation

Evisceration was carried out and hot carcass weight was taken on the slaughter floor within ten minutes post-mortem. The carcass was chilled for twenty-four hours at 3°C-4°C Cusing cold room and reweighed to get chilled carcass weight, allowed to thaw for twelve hours at room temperature and dimensional measurements: carcass length, length of hindquarters, length of fore-quarters, and depth of chest according to the procedures of De Boer *et al.* [7]. Dissection of carcasses commenced after thawing. The left sides were separated as outlined by Fisher and De Boer [7]. The lengths of the pistol were obtained following quartering of the carcass side.

### 2.6. Dissection of Muscles

The abdominal muscles were freed where they join the pelvic limb and the side was cut along the edge of *Iliocostalis lumborum* muscle through the ribs to the earlier cut between the 5<sup>th</sup> and 6<sup>th</sup> thoracic vertebrae [9] and then the cut was continued along the caudal edge of the 5<sup>th</sup> rib. This divided the side into a pistol hindquarter and a pistol forequarter. The right pistol hindquarter and pistol forequarter were weighed and dissected into individual muscles (*semimembranosus*, *semitendinosus*, *bicepsfemoris*, *adductor*, *gluteus medius*, *deltoideus*, *supraspinatus*, *infraspinatus*, *subscapularis*, and *deltodius*), bones and fat following the procedures of Robelin and Geay [10]. Values were expressed as a percentage of the pistol weight.

### 2.7. Derived Indexes

Body measurements to liveweight (cm/100g), hot carcass

weight to carcass measurements (cm/100g), and the proportion of individual muscles to pistol weight were recorded based on standard procedure [11]. Also proportion of bone distribution to pistol weight, Proportion of fat distribution to pistol weight, relationship between hot carcass weight and percentage proportion of individual muscles were obtained according to Purchas *et al.* [12].

## 2.8. Statistical Analyses

Data were statistically analyzed using one-way analysis of variance, the procedure of the Statistical Analysis System [13]. The LSD procedure was applied for mean comparison at 5%. Simple correlation coefficients of hot carcass weight and proportional weight of tissues of the proximal hindquarter and proximal forequarter of the carcasses were carried out using Pearson's correlations.

## 3. Results and Discussion

### 3.1. Body Measurements

Linear measurements of Balami, Ouda, Yakansa and West African Dwarf rams intensively fattened are shown in Table 1. There were statistical variations ( $P < 0.05$ ) in all the parameters measured except liveweight and chest width that

ranged from 36.39–37.00kg and 17.75–19.25cm. BA (115.75±4.25, 70.50±5.57 and 92.00±2.94cm) and UD (110.25±1.89, 69.00±2.16 and 84.00±1.41cm) had significantly higher values than YK (96.50±3.87, 60.75±3.30 and 76.75±0.96cm) and WAD (86.50±5.20, 53.25±5.32 and 75.25±3.30cm) for body length wither height and heart girth and the values for YK were greater than WAD rams, respectively. BA had a significant pelvic chest than UD (80.25±5.80) and WAD (61.00±2.94), which were not different ( $P > 0.05$ ). When expressed relative to liveweight, BA and WAD rams differed significantly for all values, whereas UD (84.00±1.41 and 80.25±5.80) had significantly higher values for heart girth and pelvis height than WAD (75.25±3.30 and 61.00±2.94). The effect of slaughter weight on carcass measurements indicated that all carcass measurements increased ( $P < 0.05$ ) with the increase of slaughter weight. The slaughter weight in the present study reflects commercial practices and maintains good comparability with other breeds. Lavvaf *et al.* [14] observed that chest girth and rump width were suitable factors for predicting hot carcass weight in each of Afshari and Zandi sheep, respectively. Also, body measurements are important data sources in terms of reflecting the breed standards [15], and giving information about the morphological structure and development ability of the animals.

**Table 1.** Morphometric Measurements and relative measurements to liveweight of four breeds of rams intensively fattened.

Parameters	BA	UD	YK	WAD
Live weight (kg)	36.59±1.83	37.00±0.82	36.00±1.83	36.39±2.38
Measurements (cm)				
Body length	115.75±4.25 <sup>a</sup>	110.25±1.89 <sup>a</sup>	96.50±3.87 <sup>b</sup>	86.50±5.20 <sup>c</sup>
Wither height	70.50±5.57 <sup>a</sup>	69.00±2.16 <sup>a</sup>	60.75±3.30 <sup>b</sup>	53.25±5.32 <sup>c</sup>
Heart girth	92.00±2.94 <sup>a</sup>	84.00±1.41 <sup>a</sup>	76.75±0.96 <sup>c</sup>	75.25±3.30 <sup>c</sup>
Pelvis height	88.00±2.16 <sup>a</sup>	80.25±5.80 <sup>b</sup>	71.50±2.65 <sup>c</sup>	61.00±2.94 <sup>d</sup>
Chest width	18.75±0.50	19.25±0.50	17.75±1.71	17.75±3.30
Chest depth	33.00±2.58 <sup>a</sup>	33.00±3.74 <sup>a</sup>	29.50±2.65 <sup>ab</sup>	25.75±0.96 <sup>b</sup>
Measurements relative to live weight (cm/100g)				
Body length	3.17±0.16 <sup>a</sup>	2.98±0.11 <sup>ab</sup>	2.77±0.23 <sup>b</sup>	2.76±0.26 <sup>b</sup>
Wither height	1.93±0.09 <sup>a</sup>	1.87±0.03 <sup>ab</sup>	1.74±0.16 <sup>ab</sup>	1.70±0.20 <sup>b</sup>
Heart girth	2.52±0.07 <sup>a</sup>	2.27±0.06 <sup>ab</sup>	2.20±0.12 <sup>bc</sup>	2.40±0.17 <sup>c</sup>
Pelvis height	2.41±0.09 <sup>a</sup>	2.17±0.15 <sup>b</sup>	2.05±0.17 <sup>bc</sup>	1.94±0.14 <sup>c</sup>
Chest width	0.51±0.04	0.52±0.02	0.51±0.07	0.57±0.11
Chest depth	0.09±0.07	0.89±0.10	0.85±0.09	0.82±0.08

Means in the same row with different superscripts are significantly different ( $P < 0.05$ ). BA: Balami, UD: Ouda, YK: Yankasa, WAD: West African Dwarf

### 3.2. Carcass Measurements

Table 2 summarized the carcass measurements and distribution of tissues within the proximal hindquarters and forequarters. The proportion of hot carcass weight was 18.72±1.85, 18.25±0.87, 17.72±0.17 and 17.72±1.26 for BA, UD, YK, and WAD, respectively. Pistola weights of the hindquarter and forequarter were significantly greater for BA (74.00±4.36; 61.67±7.10) and UD (68.00±6.25; 60.67±2.60) than YK (62.67±4.16; 54.67±7.23) which in turn was greater than WAD (50.00±3.61; 49.00±3.61). When expressed relative to hot carcass weight, carcass length, carcass depth,

hindquarters, forequarters, and Pistola lengths did not differ ( $P > 0.05$ ) between the breeds but the lowest were recorded ( $P < 0.05$ ) for WAD. All the four breeds differed significantly for lengths of hindquarters and forequarters relative to carcass weight, which was greatest for BA (0.47±0.04; 0.33±0.01) and least for WAD (0.34±0.02; 0.28±0.01). Carcass measurements and indexes are used as indicators of carcass conformation and size [16]. The results of the current study indicate a significant effect of breed on carcass measurements and conformation indexes, which could be explained by breed specific morphologic characteristics of

BA rams.

**Table 2.** Hot carcass weight and carcass measurements of four breeds of rams intensively fattened.

Parameters	BA	UD	YK	WAD
Hot Carcass Weight	18.72±1.85	18.25±0.87	17.72±0.17	17.72±1.26
Carcass Measurements (cm)				
Carcass Length	75.75±4.79 <sup>a</sup>	75.50±4.66 <sup>a</sup>	59.00±7.79 <sup>b</sup>	60.50±3.11 <sup>b</sup>
Carcass Depth	24.25±0.96	24.50±0.58	24.75±3.30	24.25±4.03
Hindquarter Length	88.00±2.16 <sup>a</sup>	80.25±5.80 <sup>b</sup>	71.50±2.65 <sup>c</sup>	61.00±2.94 <sup>d</sup>
Pistola Hindquarter Length	74.00±4.36 <sup>a</sup>	68.00±6.25 <sup>ab</sup>	62.67±4.16 <sup>b</sup>	50.00±3.61 <sup>c</sup>
Forequarter Length	70.30±6.81 <sup>a</sup>	69.00±2.65 <sup>a</sup>	64.00±8.55 <sup>ab</sup>	55.33±4.04
Pistola Forequarter Length	61.67±7.10 <sup>a</sup>	60.67±25.89 <sup>a</sup>	54.67±7.23 <sup>ab</sup>	49.00±3.61 <sup>b</sup>
Carcass Measurements (cm/100g carcass)				
Carcass Length	0.41±0.02 <sup>a</sup>	0.42±0.02 <sup>a</sup>	0.34±0.04 <sup>b</sup>	0.39±0.05 <sup>ab</sup>
Carcass Depth	0.13±0.01	0.14±0.01	0.14±0.02	0.15±0.01
Hindquarter Length	0.47±0.04 <sup>a</sup>	0.44±0.02 <sup>ab</sup>	0.40±0.03 <sup>b</sup>	0.34±0.01 <sup>c</sup>
Pistola Hindquarter Length	0.40±0.04 <sup>a</sup>	0.37±0.02 <sup>ab</sup>	0.34±0.02 <sup>b</sup>	0.28±0.01 <sup>c</sup>
Forequarter Length	0.38±0.02 <sup>a</sup>	0.38±0.04 <sup>a</sup>	0.35±0.03 <sup>ab</sup>	0.31±0.02 <sup>b</sup>
Pistola Forequarter Length	0.33±0.01 <sup>a</sup>	0.33±0.04 <sup>a</sup>	0.30±0.03 <sup>ab</sup>	0.28±0.01 <sup>b</sup>

Means in the same row with different superscripts are significantly different (P<0.05). BA: Balami, UD: Ouda, YK: Yankasa, WAD: West African Dwarf

**3.3. Meat to Bone Ratio**

Weight to length ratio has been proposed as an objective measure of the carcass index. This index describes the amount of meat and fat which a carcass carries relative to its length and affords a quick and accurate description of its meatiness [17]. Proportions of tissues in the carcass of the four breeds of rams intensively fattened are shown in Table 3. The meat to bone ratio as determined by dissection of the proximal hindquarter and proximal forequarter of the left half of the carcass were 8.92 and 12.90 in BA and 5.81 and 12.64 in UD, respectively and the corresponding ratio for YK and WAD were 5.46; 7.92 and 4.88; 10.10, respectively. BA had the highest (P<0.05) meat to bone ratio due to its lowest fat proportion. UD, YK, and WAD yielded similar meat to the bone ratio (P>0.05) while BA, UD, and WAD breeds had similar meat to fat ratios. Differences in carcass were

primarily due to the distribution of fat and to variation in the thickness of muscles but not differences in muscle distribution. The sequence of growth illustrates the importance of each fat depot in animals and the market value of the carcass [18]. Cloete *et al.* [19] observed differences in the fat deposition of mutton breeds which suggests that different breeds would deposit fat at different levels. Early matured breeds would render a fatter carcass at slaughter, after a constant number of days in the feed [20], which in turn would affect the value of the carcass. It is therefore important to determine the maturation rate of the major carcass tissues of various breeds of sheep to accurately predict a slaughter weight for ideal grading of the carcass. Using a commercial dissection procedure, Colomer-Rocheret *al.* [21] found that hindquarter cuts had more muscle and a higher muscle-to-bone ratio than those in forequarter cuts.

**Table 3.** Proportional Percentage of Muscle, Bone and Fat to the Pistola Weight of the Hindlegs and Forelegs of four breeds of rams intensively fattened (±STD).

Breeds	BA	UD	YK	WAD
Pistola Weight of Hindleg	4.72±0.58 <sup>a</sup>	5.35±0.19 <sup>ab</sup>	4.04±1.09 <sup>ab</sup>	4.67±0.35 <sup>b</sup>
Pistola Weight of Foreleg	2.26±0.32	2.81±0.21	2.49±0.50	2.64±0.45
Muscle distribution (g/kg)				
Pistola Muscles of Hindquarter	82.48±0.01 <sup>a</sup>	82.44±3.74 <sup>a</sup>	72.88±4.11 <sup>b</sup>	77.71±0.89 <sup>ab</sup>
Pistola Muscles of Foreleg	82.18±0.01 <sup>a</sup>	76.97±3.09 <sup>ab</sup>	73.79±4.97 <sup>b</sup>	72.22±6.53 <sup>b</sup>
Bone distribution (g/kg)				
Pistola Bones of Hindquarter	9.03±0.01 <sup>b</sup>	12.28±3.66 <sup>ab</sup>	16.68±4.22 <sup>a</sup>	15.19±3.08 <sup>a</sup>
Pistola Bones of Foreleg	9.43±0.02 <sup>b</sup>	15.33±2.74 <sup>a</sup>	14.15±1.62 <sup>ab</sup>	13.08±4.01 <sup>ab</sup>
Fat distribution (g/kg)				
Pistola Fat of Hindquarter	7.12±0.06 <sup>b</sup>	5.24±1.38 <sup>b</sup>	8.23±3.04 <sup>ab</sup>	11.49±1.89 <sup>a</sup>
Pistola Fat of Foreleg	5.64±0.03	5.37±1.87	6.68±0.37	7.61±2.18
Meat: Bone	8.92±0.01 <sup>a</sup>	5.81±0.04 <sup>b</sup>	5.46±1.01 <sup>b</sup>	4.88±0.97 <sup>b</sup>
Meat: Fat	12.90±0.03 <sup>a</sup>	12.64±0.28 <sup>a</sup>	7.92±2.52 <sup>b</sup>	10.10±2.00 <sup>ab</sup>

Means in the same row with different superscripts are significantly different (P<0.05). BA: Balami, UD: Ouda, YK: Yankasa, WAD: West African Dwarf

**3.4. Muscle Measurements of the Pistola Hindquarters and Forequarters**

The distribution of muscles in intensively fattened Balami, Ouda, Yankasa and West African Dwarf rams is shown in

Table 4. The highest value (P<0.05) of *semimembranosus* was recorded for BA (0.54±0.02) and while the least was recorded for WAD (0.47±0.03). The proportional values measured values for Ouda (0.51±0.08) and Yankasa

(0.45±0.01) in the *semimembranosus* muscle were higher ( $P<0.05$ ) than Balami (0.39±0.03) and West African Dwarf rams (0.38±0.02). Results showed that Ouda (0.38±0.03) had the highest *Adductor* muscle (%), followed by Yankasa (0.37±0.01%) while Balami (0.33±0.01) and WAD breeds (0.32±0.04) showed the lowest. The percentage proportion of *Gluteus Medius* muscle was higher in Ouda (0.14±0.01) when compared to Yankasa (0.13±0.01), but not significant when compared to Balami (0.13±0.01) and WAD (0.12±0.01). The proportional weight of *gluteus medius* was lower for YK (0.15±0.01) and WAD (0.15±0.02) than BA (0.18±0.01) and UD (0.17±0.01), which were similar ( $P>0.05$ ). The weight of *deep pectoralis* was higher for BA (0.63±0.06) than UD (0.42±0.11), YK (0.38±) and WAD (0.42±0.17), which were not significant to one another ( $P>0.05$ ). The weights of *infraspinatus* (0.23±0.01), *supraspinatus* (0.18±0.05), *deltoideus* (0.30±0.02) and *subscapularis* (0.24±0.08) were significantly greater for BA. The corresponding weights of *infraspinatus* (0.18±0.04), *supraspinatus* (0.13±0.01), *deltoideus* (0.25±0.02), and *subscapularis* (0.27±0.10) were not significantly different ( $P>0.05$ ) for WAD rams. The relative weight of the *semimembranosus* was significantly higher for UD (0.51±0.08) and YK (0.45±0.01) than for BA (0.39±0.03)

and WAD (0.38±0.02). The relative weight of the *Semitendinosus* muscle (0.40±0.04), *Adductor* (0.38±0.03), *Gluteus medius* (0.14±0.01) and *Biceps femoris* (0.34±0.04) were significantly higher for Ouda than the values recorded for both BA and YK. With increasing slaughter weight, muscle weight increased significantly for *semimembranosus*, *semitendinosus*, *gluteus medius*, *biceps femoris*, *deep pectoralis*, *infraspinatus*, *supraspinatus*, *deltoideus* and *subscapularis*. All relative measurements decreased significantly for BA except for *deep pectoralis*, *infraspinatus*, and *supraspinatus*, which did not differ ( $P>0.05$ ) between the slaughter weights. The greater muscle of BA compared to the UD, YK and WAD breeds reflected the fleshiness and carcass size. An increase in carcass weight increased the values of all carcass measurements but when expressed relative to carcass weight they were reduced, indicating greater carcass compactness. The relatively small differences in tissue distribution together with the significant differences in the proportion of muscle in the weight suggest that carcass weight is a reliable indicator of economic traits. Lodge [22] suggested that the superior performance of local sheep is due to the results that they have the potential to respond to better management.

**Table 4.** Hot carcass weight and percentage proportion of muscles of four breeds of rams intensively fattened.

Breeds	BA	UD	YK	WAD
Hot Carcass Weight	18.72±1.85	18.25±0.87	17.72±0.17	16.82±1.26
Muscle Measurements (g)				
Semimembranosus	0.54±0.02 <sup>ab</sup>	0.59±0.10 <sup>a</sup>	0.49±0.01 <sup>b</sup>	0.47±0.03 <sup>b</sup>
Semitendinosus	0.49±0.14 <sup>a</sup>	0.46±0.02 <sup>ab</sup>	0.37±0.02 <sup>ab</sup>	0.35±0.06 <sup>b</sup>
Adductor	0.47±0.06	0.45±0.03	0.40±0.01	0.40±0.05
GluteusMedius	0.18±0.01 <sup>a</sup>	0.17±0.01 <sup>ab</sup>	0.15±0.01 <sup>c</sup>	0.15±0.02 <sup>bc</sup>
Bicepsfemoris	0.36±0.01 <sup>a</sup>	0.39±0.06 <sup>a</sup>	0.25±0.05 <sup>b</sup>	0.22±0.06 <sup>b</sup>
Deepectoralis	0.63±0.06 <sup>a</sup>	0.42±0.11 <sup>b</sup>	0.38±0.02 <sup>b</sup>	0.42±0.17 <sup>b</sup>
Infraspinatus	0.23±0.01 <sup>a</sup>	0.19±0.01 <sup>b</sup>	0.18±0.01 <sup>b</sup>	0.18±0.04 <sup>b</sup>
Supraspinatus	0.18±0.05 <sup>a</sup>	0.14±0.01 <sup>b</sup>	0.14±0.01 <sup>b</sup>	0.13±0.01 <sup>b</sup>
Deltoideus	0.30±0.02 <sup>a</sup>	0.27±0.01 <sup>ab</sup>	0.28±0.02 <sup>ab</sup>	0.25±0.02 <sup>b</sup>
Subscaplaris	0.24±0.08 <sup>ab</sup>	0.17±0.09 <sup>b</sup>	0.31±0.01 <sup>a</sup>	0.27±0.10 <sup>ab</sup>
Percentage proportion of muscle to HCW				
Semimembranosus	0.39±0.03 <sup>bc</sup>	0.51±0.08 <sup>a</sup>	0.45±0.01 <sup>ab</sup>	0.38±0.02 <sup>c</sup>
Semitendinosus	0.34±0.06 <sup>ab</sup>	0.40±0.04 <sup>a</sup>	0.34±0.02 <sup>ab</sup>	0.28±0.04 <sup>b</sup>
Adductor	0.33±0.01 <sup>bc</sup>	0.38±0.03 <sup>a</sup>	0.37±0.01 <sup>ab</sup>	0.32±0.04 <sup>c</sup>
GluteusMedius	0.13±0.01 <sup>bc</sup>	0.14±0.01 <sup>a</sup>	0.13±0.00 <sup>b</sup>	0.12±0.01 <sup>c</sup>
Bicepsfemoris	0.25±0.01 <sup>b</sup>	0.34±0.04 <sup>a</sup>	0.24±0.05 <sup>bc</sup>	0.18±0.04 <sup>c</sup>
Deepectoralis	0.45±0.09	0.37±0.11	0.35±0.01	0.34±0.13
Infraspinatus	0.17±0.01	0.16±0.01	0.17±0.01	0.14±0.03
Supraspinatus	0.13±0.04	0.12±0.00	0.13±0.01	0.11±0.01
Deltoideus	0.21±0.01 <sup>bc</sup>	0.23±0.00 <sup>b</sup>	0.26±0.02 <sup>a</sup>	0.20±0.02 <sup>c</sup>
Subscaplaris	0.17±0.04 <sup>b</sup>	0.22±0.06 <sup>ab</sup>	0.28±0.01 <sup>a</sup>	0.27±0.01 <sup>a</sup>

Means in the same row with different superscripts are significantly different ( $P<0.05$ ). BA: Balami, UD: Ouda, YK: Yankasa, WAD: West African Dwarf

### 3.5. Relationship Between Hot Carcass Weight and Muscle Distribution

Significant positive correlations ( $P<0.05$ ) were obtained between hot carcass weight and *Semitendinosus* muscle (0.89) and *Supraspinatus* muscle (0.56), whereas a high

negative correlation ( $P<0.05$ ) was obtained with *Subscapularis* muscle (0.34). Significant positive correlations were obtained between *Semitendinosus* muscle and *Biceps femoris* muscle (0.58), *Adductor* muscle (0.88), *Gluteus medius* (0.83), *Infraspinatus* (0.67) and *Deltodius* (0.66),

whereas significant negative correlations were obtained with *Subscapularis* muscle and *Biceps femoris* muscle (0.50), *Semimembranosus* muscle (0.37), *Adductor* muscle (0.97), *Deep pectoralis* muscle (0.54) and *Supraspinatus* muscle (0.73) proportions. Correlation coefficients between *Semimembranosus* muscle and *Biceps femoris* muscle (0.84), *Adductor* muscle (0.68) and *Gluteus medius* (0.64) were in the same direction of those obtained from *Biceps femoris* muscle with *Adductor* muscle (0.61) and *Gluteus medius* (0.71) value. Correlations between the muscles of the carcass

appear more dependent upon hindlegs and forelegs rather than an absolute hot carcass weight. This is evidenced by the fact that the highest correlations were seen between the muscles of the foreleg (*supraspinatus* and *infraspinatus*, *Deep pectoralis* and *Deltoideus* muscles) and the hindleg (*semimembranosus* and *semitendinosus*, *Adductor* and *Gluteus medius* muscles). The relationship between hot carcass weight and the distribution of muscles suggests that an increase in one could lead to a corresponding increase in the other muscles [23, 24].

**Table 5.** Pearson Correlation coefficients between hot carcass weight and percentage proportion of muscles of carcasses of Balami, Ouda, Yankasa and West African Dwarf rams intensively fattened.

	HCW	ST	SM	BF	ADD	GM	DP	IF	SS	DT	SCP
HCW	1	0.89*	0.52 <sup>ns</sup>	0.29 <sup>ns</sup>	0.77 <sup>ns</sup>	0.67 <sup>ns</sup>	0.48 <sup>ns</sup>	0.39 <sup>ns</sup>	0.56*	0.32 <sup>ns</sup>	-0.34 <sup>ns</sup>
ST		1	0.42 <sup>ns</sup>	0.58*	0.88***	0.83***	0.38 <sup>ns</sup>	0.67**	0.79 <sup>ns</sup>	0.66**	0.80 <sup>ns</sup>
SM			1	0.84***	0.68**	0.64**	0.42 <sup>ns</sup>	0.43 <sup>ns</sup>	0.33 <sup>ns</sup>	0.29 <sup>ns</sup>	-0.37 <sup>ns</sup>
BF				1	0.61**	0.71**	0.37 <sup>ns</sup>	0.36 <sup>ns</sup>	0.32 <sup>ns</sup>	0.41 <sup>ns</sup>	-0.50*
ADD					1	0.88***	0.54*	0.67**	0.43 <sup>ns</sup>	0.63**	-0.97**
GM						1	0.55*	0.75***	0.39 <sup>ns</sup>	0.54*	-0.60 <sup>ns</sup>
DP							1	0.80***	0.64**	0.46 <sup>ns</sup>	-0.54 <sup>ns</sup>
IF								1	0.57*	0.78***	0.49 <sup>ns</sup>
SS									1	0.37	-0.73 <sup>ns</sup>
DT										1	0.42 <sup>ns</sup>
SCP											1

HCW: Hot carcass Weight, ST: Semitendinosus muscle, SM; Semimembranosus muscle, BF: Biceps femoris, ADD; Adductor, GM: Gluteus medius, DP: Deep pectoralis, IF: Infraspinatus, SS: Supraspinatus, DT: Deltoideus, SCP: Subscapularis, Significance levels: \*\*\*P<0.001, \*\*P<0.01, \*P<0.05

## 4. Conclusions

Differences between breeds were observed in the morphology of the carcass and muscles of four breeds of rams intensively fattened. Yankasa and West African Dwarf breeds of rams had lower carcass measurements than Balami and Ouda breeds. The combination of morphological traits and distribution of muscles found in the carcasses for the Balami breeds of rams in this study indicated that this breed would most likely produce the best carcass under similar agricultural conditions. The study also concludes that the measurements of the muscles of the hindlegs and forelegs could be used satisfactorily for prediction of hot carcass weight.

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## Conflict of Interest

The authors declare that they have no conflict of interest.

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